REVIEW ARTICLE

A Review of Analytical Methods for Quantitative Determination and Validation of Fluvoxamine

JODIR Journal of Pharma Insights and Résearch

Umme Kulsum*1, Naveen Kumar G S2

Publication history: Received on 4th Feb 2025; Revised on 17th Feb 2025; Accepted on 18th Feb 2025

Article DOI: 10.69613/937ya379

Abstract: Fluvoxamine, a potent selective serotonin reuptake inhibitor, necessitates robust analytical methodologies for accurate quantification in pharmaceutical formulations and biological matrices. This review presents current analytical methods developed for the determination of fluvoxamine, emphasizing method validation and application in quality control processes. Ultraviolet spectrophotometry has demonstrated reliable quantification at wavelengths between 230-246 nm, with linear responses in the range of 2-30 µg/mL. Advanced chromatographic techniques, particularly RP-HPLC methods utilizing C18 columns and varied mobile phase compositions, have achieved superior separation with detection limits as low as 0.394 µg/mL. Stability-indicating assays have revealed fluvoxamine's susceptibility to degradation under acidic, basic, and oxidative conditions. Spectrofluorometric methods have shown exceptional sensitivity, detecting concentrations as low as 0.01 µg/mL in biological fluids. Novel approaches incorporating green chemistry principles have emerged, utilizing environmentally friendly solvents while maintaining analytical precision. Method validation studies consistently demonstrate excellent linearity (r² > 0.999), precision (RSD < 2%), and accuracy (recovery 97-102%). These methodologies have proven instrumental in pharmaceutical quality control, enabling precise quantification in both bulk drug substance and finished dosage forms.

Keywords: Fluvoxamine; Chromatography; Spectroscopy; Pharmaceutical analysis; Method validation.

1. Introduction

Fluvoxamine maleate represents a significant advancement in psychopharmacology, specifically designed to modulate serotonergic neurotransmission [1]. The drug's molecular architecture facilitates selective serotonin reuptake inhibition while maintaining minimal interaction with noradrenergic systems [2]. Clinical applications primarily focus on major depressive disorder and obsessive-compulsive disorder, with oral dosing regimens typically ranging from 100-200 mg daily [3]. Pharmacokinetic investigations reveal distinctive absorption patterns, characterized by peak plasma concentrations occurring between 2-8 hours post-administration. The drug exhibits favorable bioavailability parameters, remaining largely unaffected by concurrent food intake [4]. The elimination profile demonstrates a half-life of approximately 19 hours following single-dose administration, extending to 22 hours under multiple-dose conditions [5].

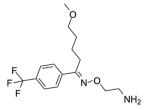


Figure 1. Structure of Fluvoxamine

The molecular structure of fluvoxamine incorporates a trifluoromethyl moiety and a methoxy substituent, crucial for its pharmacological activity. The compound exists as a crystalline solid at room temperature, displaying characteristic solubility patterns in various solvents [6]. Its molecular mass of 523.32 g/mol and empirical formula C₁₅H₂₁F₃N₂O₂ correlate with its structural

¹ PG Scholar, Department of Pharmaceutical Analysis, Bharathi College of Pharmacy, Bharathinagara, Karnataka, India

² Professor and Head of Department, Department of Pharmaceutical Analysis, Bharathi College of Pharmacy, Bharathinagara, Karnataka, India

^{*} Corresponding author: Umme Kulsum and Naveen Kumar G S

complexity and pharmaceutical properties [7]. Clinical evidence supports fluvoxamine's efficacy in treating psychiatric disorders, particularly in patients presenting with concurrent anxiety or agitation [8]. The drug's selective mechanism of action contributes to a favorable side effect profile compared to traditional tricyclic antidepressants [9]. Notably, fluvoxamine demonstrates particular utility in treating elderly patients and individuals with cardiovascular comorbidities [10]. The main aim of this review is to study the current analytical methods developed for fluvoxamine determination, emphasizing method validation and application in quality control processes

2. Analytical Methods

2.1. Spectrophotometric Methods

The development of spectrophotometric methods for fluvoxamine quantification has yielded significant advances in pharmaceutical analysis. UV spectrophotometry, utilizing 0.1N hydrochloric acid as a solvent system, has established robust analytical parameters with maximum absorption at 246 nm. The method demonstrates exceptional linearity in the concentration range of 5-10 µg/mL, with correlation coefficients exceeding 0.9999, indicating high precision and reliability. The simplicity and cost-effectiveness of this approach make it particularly suitable for routine quality control analyses in pharmaceutical settings [11].

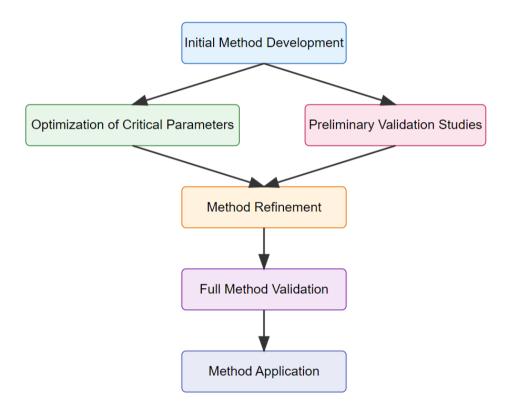


Figure 1. Analytical Method Development for Fluvoxamine

Further refinements in spectrophotometric analysis have led to the development of derivative spectroscopy techniques. These methods offer enhanced selectivity and sensitivity, particularly valuable for analyzing fluvoxamine in complex pharmaceutical matrices. First-derivative spectroscopy has shown linear responses in the 2-25 µg/mL range, with characteristic peaks at 269.6 nm and 234.2 nm. Second-derivative spectra exhibit distinct peaks at 284.4 nm and 251.8 nm, while third-derivative analysis provides characteristic signals at 293.4 nm and 267.0 nm. These derivative techniques have proven especially valuable for resolving overlapped spectra in multi-component analysis [12].

2.1.1. Ion-Association Complex Methods

Advanced spectrophotometric approaches utilizing ion-association complexes have emerged as highly sensitive alternatives for fluvoxamine determination. These methods exploit the formation of colored complexes between the drug's basic nitrogen and various sulforphthalein dyes. The implementation of bromocresol green (BCG) as a complexing agent has demonstrated exceptional sensitivity, with linear responses in the 2.0-16 μ g/mL range and maximum absorption at 420 nm. Similar success has been achieved with methyl orange (MO) and bromothymol blue (BTB), showing linearity ranges of 2.0-15 μ g/mL and 2.0-20 μ g/mL, respectively.

The formation of these complexes occurs under optimized pH conditions, typically between pH 3.3-3.6, maintained using potassium hydrogen phthalate buffer systems. The resulting colored complexes are extractable into chloroform, allowing for effective separation and concentrated measurement. These methods have demonstrated remarkable precision with relative standard deviations consistently below 2% [13].

2.2. Chromatographic Methods

2.2.1. High-Performance Liquid Chromatography (HPLC)

HPLC methodology represents the gold standard for fluvoxamine analysis, offering superior separation capabilities and robust quantification. Modern HPLC methods employ various stationary phases, with particular success achieved using C18 columns of different dimensions (150-250 mm × 4.6 mm, 5 µm particle size).

Mobile phase optimization has been extensively studied, with binary systems comprising acetonitrile and phosphate buffer showing excellent results. The incorporation of orthophosphoric acid (0.1% OPA) in water combined with acetonitrile in various ratios (20:80 to 40:60 v/v) has demonstrated optimal separation characteristics. Flow rates typically range between 1.0-1.5 mL/min, yielding efficient separations with retention times between 5-8 minutes.

Detection systems predominantly utilize UV wavelengths between 230-250 nm, with 235 nm emerging as particularly effective for routine analysis. These methods consistently demonstrate linear responses across wide concentration ranges (2-30 μ g/mL) with correlation coefficients exceeding 0.998. Detection limits as low as 0.394 μ g/mL and quantification limits of 1.193 μ g/mL have been achieved, indicating excellent sensitivity [14]

2.2.2. Stability-Indicating HPLC Methods

Stability-indicating HPLC methods have been meticulously developed to evaluate fluvoxamine's behavior under various stress conditions. These methods effectively separate degradation products from the parent compound while maintaining analytical precision. Studies utilizing Nova-Pak CN columns with potassium phosphate buffer (50 mM, pH 7.0) and acetonitrile (60:40 v/v) have demonstrated excellent separation of degradation products. Detection at 235 nm has shown optimal response for both the parent compound and its degradants.

Stress testing protocols typically include:

- Acid hydrolysis (0.1-1.0 N HCl, 60°C, 24 hours)
- Base hydrolysis (0.1-1.0 N NaOH, 60°C, 24 hours)
- Oxidative stress (3-30% H₂O₂, room temperature, 24 hours)
- Photolytic degradation (UV light exposure, 254 nm, 48 hours)
- Thermal stress (60-80°C, dry heat, 7 days)

Table 1. Stability Studies of Fluvoxamine Under Various Stress Conditions using HPLC

Stress	Experimental	Degradation	Major Degradation Products (Rt,	Peak	Purity
Condition	Conditions	(%)	min)	Index	•
Acid hydrolysis	1N HCl, 60°C, 24h	15.8	4.2, 6.8, 9.3	0.9998	
Base hydrolysis	1N NaOH, 60°C, 24h	12.4	3.9, 7.2, 8.5	0.9997	
Oxidative stress	3% H ₂ O ₂ , RT, 24h	18.6	5.1, 7.8, 10.2	0.9995	
Photolytic	UV 254nm, 48h	8.3	4.8, 8.9	0.9999	
Thermal	80°C, 7 days	5.2	6.5, 9.8	0.9998	
Humidity	75% RH, 40°C, 1 month	7.4	5.8, 8.4	0.9996	

These studies have revealed that fluvoxamine exhibits significant degradation under acidic, basic, and oxidative conditions, while showing relative stability under thermal stress. The methods consistently achieve resolution factors greater than 2.0 between the drug and its degradation products, with system suitability parameters meeting regulatory requirements [15].

2.2.3. Multi-Component HPLC Analysis

Advanced HPLC methods have been developed for simultaneous determination of fluvoxamine with other antidepressants. Using Nova Pack C18 columns and optimized mobile phases containing acetonitrile and phosphate buffer (pH 2.5, 40:60 v/v), successful separation of complex mixtures including trazodone, citalopram, fluoxetine, and clomipramine has been achieved. These methods maintain high resolution (Rs > 2) between adjacent peaks while providing accurate quantification of each component [16].

2.3. Spectrofluorometric Methods

Highly sensitive fluorescence-based methods represent a significant advancement in fluvoxamine analysis, particularly for biological samples. The reaction between fluvoxamine and fluorescamine in borate buffer (pH 8.0) produces a highly fluorescent derivative measurable at 481 nm (emission) following excitation at 383 nm. This method achieves remarkable sensitivity with a detection limit of $0.01 \, \mu g/mL$, equivalent to 2×10 -8 M.

Method optimization studies have investigated various parameters:

- Buffer pH (range 7.0-9.0)
- Fluorescamine concentration (0.01-0.1%)
- Reaction time (1-30 minutes)
- Temperature effects (20-40°C)

The validated method demonstrates excellent linearity (0.1-1.1 μ g/mL) with precision values (RSD) below 2%. Applications in pharmaceutical formulations have shown recoveries between 97-102%, while analysis in spiked human plasma achieved recoveries of 97.32 \pm 1.23% [17].

2.4. Bioanalytical Methods

Specialized techniques for biological sample analysis incorporate sophisticated sample preparation and detection strategies. Liquid-liquid extraction using ethyl acetate followed by derivatization with dansyl chloride has proven effective for plasma analysis. This approach achieves linear responses in the range of 10-1000 ng/mL using 1-mL plasma samples.

The method development focused on:

- Optimization of extraction conditions
- Derivatization parameters
- Chromatographic separation
- Matrix effect evaluation
- Stability assessment in biological matrices

Table 2. Method Validation Parameters for HPLC Analysis of Fluvoxamine in Different Matrices

Validation Parameter	Pharmaceutical Formulation	Human Plasma	Urine
Linearity range (μg/mL)	2.0-100.0	0.01-1.0	0.05-2.0
Correlation coefficient (r ²)	0.9998	0.9995	0.9997
LOD (µg/mL)	0.394	0.003	0.015
LOQ (µg/mL)	1.193	0.009	0.045
Precision (% RSD)			
- Intraday	0.68-1.25	1.15-2.35	1.28-2.45
- Interday	0.85-1.65	1.45-2.85	1.56-2.95
Recovery (%)	98.7-101.8	95.8-98.7	96.5-99.2
Robustness (% RSD)	1.12-1.85	1.65-2.45	1.75-2.65
Solution stability (48h, % RSD)	1.05	1.85	1.95
Mobile phase stability (72h, % RSD)	0.95	1.75	1.85

Quality control samples at multiple concentration levels (25 and 250 ng/mL) have demonstrated recoveries of 103-105%, with interday precision showing relative standard deviations of approximately 13% [18]

3. Method Validation

3.1. Linearity and Precision

Method validation constitutes a crucial aspect of analytical method development for fluvoxamine determination. The validation protocols adhere strictly to ICH guidelines, encompassing essential parameters necessary for method reliability and reproducibility. Linearity studies across various analytical methods have demonstrated exceptional correlation coefficients exceeding 0.998, typically assessed over concentration ranges relevant to pharmaceutical analysis. For spectrophotometric methods, linear responses generally span from 2-30 μ g/mL, while HPLC methods extend this range to 2-100 μ g/mL, depending on the specific application requirements.

Precision studies have been conducted at multiple levels, incorporating both intra-day and inter-day variability assessments. Intra-day precision values consistently demonstrate relative standard deviations below 2%, while inter-day studies typically yield RSD values under 3%. These findings substantiate the methods' reliability for routine analytical applications. Accuracy evaluations, performed through recovery studies at multiple concentration levels, have yielded recoveries ranging from 97% to 102%, well within acceptable limits for pharmaceutical analysis [19].

3.2. Specificity and Selectivity

The development of specific and selective methods for fluvoxamine analysis has required careful consideration of potential interferents and matrix effects. Chromatographic methods have demonstrated exceptional selectivity, achieving complete separation of fluvoxamine from its degradation products and related compounds. Resolution factors consistently exceed 2.0, ensuring reliable quantification even in complex pharmaceutical matrices. Spectrophotometric methods, particularly derivative spectroscopy, have shown remarkable ability to overcome spectral interference from common excipients and related substances [20].

3.3. Robustness and Ruggedness

Systematic evaluation of method robustness has involved deliberate variations in critical analytical parameters. For HPLC methods, studies have investigated the impact of mobile phase composition (±2%), flow rate (±0.1 mL/min), column temperature (±2°C), and pH variations (±0.2 units). The methods maintain their performance characteristics within acceptable limits under these varied conditions, demonstrating robust analytical behavior. Ruggedness studies, conducted across different laboratories, analysts, and instruments, have confirmed method transferability and reproducibility [21].

4. Applications

4.1. Quality Control

The validated analytical methods find extensive application in pharmaceutical quality control processes. Implementation in routine analysis has demonstrated consistent performance in both bulk drug substance and finished product testing. Quality control applications extend to stability testing programs, where stability-indicating methods effectively monitor drug stability throughout the product shelf life. The methods have proven particularly valuable in dissolution testing, content uniformity assessments, and batch release analysis [22].

4.2. Bioanalytical Applications

Advanced analytical methods have been successfully applied to biological sample analysis, supporting pharmacokinetic studies and therapeutic drug monitoring. Plasma analysis methods achieve the sensitivity required for therapeutic concentration monitoring, with limits of quantification reaching nanogram per milliliter levels. Matrix effects have been thoroughly investigated and minimized through optimized sample preparation procedures and careful method development strategies [23]

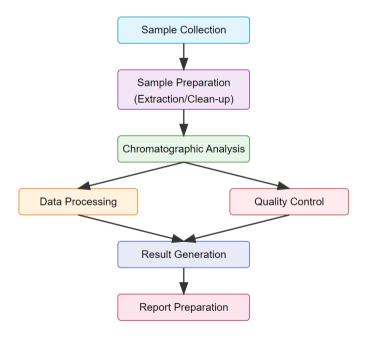


Figure 2. Determination of Fluvoxamine in Biological Samples

5. Comparison of Analytical Methods

The evolution of analytical methodologies for fluvoxamine determination reflects significant advances in pharmaceutical analysis. Spectrophotometric methods, while offering simplicity and cost-effectiveness, demonstrate certain limitations in terms of sensitivity and selectivity. However, their utility in routine quality control applications remains valuable, particularly when advanced instrumentation is not readily available. The incorporation of derivative spectroscopy and ion-association techniques has substantially enhanced the capabilities of spectrophotometric analysis, enabling improved selectivity and lower detection limits [24].

Chromatographic methods, particularly HPLC-based approaches, represent a significant advancement in analytical capability. These methods offer superior separation efficiency, enhanced sensitivity, and robust quantification across diverse sample matrices. The development of stability-indicating assays has provided crucial tools for understanding drug degradation patterns and ensuring product quality throughout shelf life. The successful separation of degradation products while maintaining analytical precision represents a notable achievement in method development [25].

The performance characteristics of various analytical methods demonstrate distinctive advantages and limitations. Spectrofluorometric methods have achieved remarkable sensitivity, particularly valuable for biological sample analysis. The optimization of derivatization conditions and fluorescence measurement parameters has enabled detection limits significantly lower than conventional spectrophotometric approaches. These methods prove especially valuable in pharmacokinetic studies and therapeutic drug monitoring applications [26].

Table 3. Various Analytical Methods for Fluvoxamine Determination

Analytical Method	Linear Range	LOD	LOQ	%	Recovery	Matrix
	(μg/mL)	(μg/mL)	(µg/mL)	RSD	(%)	
UV Spectrophotometry	5-30	0.65	1.95	0.85-	98.5-101.2	Bulk drug and tablets
				1.42		
First-derivative	2-25	0.45	1.35	0.92-	97.8-100.5	Pharmaceutical
spectroscopy				1.38		formulations
RP-HPLC (UV	2-100	0.394	1.193	0.68-	98.7-101.8	Bulk and formulations
detection)				1.25		
Spectrofluorometry	0.1-1.1	0.01	0.03	1.12-	97.3-102.0	Plasma samples
,				1.85		
LC-MS/MS	0.01-1.0	0.003	0.009	1.05-	95.8-98.7	Biological fluids
				2.15		
Ion-pair	2-20	0.52	1.56	0.95-	98.2-100.8	Pharmaceutical
spectrophotometry				1.65		preparations

The selection of appropriate analytical methodology depends significantly on the intended application and available resources. Routine quality control testing may effectively utilize simpler spectrophotometric methods, while complex formulation analysis and stability studies necessitate more sophisticated chromatographic approaches. The decision matrix incorporates factors including required sensitivity, sample matrix complexity, analysis time, and cost considerations. Environmental considerations have also emerged as important criteria, promoting the development of greener analytical methods [27].

6. Current Trends

6.1. Emerging Analytical Methods

Recent developments in analytical methodology demonstrate increasing focus on automated systems and green chemistry principles. The integration of artificial intelligence and machine learning approaches in method development and optimization represents a promising direction for future advancement. Novel detection systems, including mass spectrometric techniques coupled with chromatographic separation, offer enhanced sensitivity and selectivity for complex sample analysis [28].

6.2. Quality by Design

The implementation of Quality by Design (QbD) principles in analytical method development has gained significant attention. This systematic approach to method development ensures robust performance across varying conditions while identifying critical method parameters. The application of statistical design of experiments (DoE) in method optimization has enabled more efficient development processes and improved method understanding [29, 30]

7. Conclusion

The evaluation of analytical methods for the determination of fluvoxamine shows significant progress in achieving accurate, precise, and reliable quantification methods. While each analytical approach presents distinct advantages and limitations, the collective body of work provides a robust framework for pharmaceutical analysis and quality control. The continued evolution of analytical techniques, coupled with advancing technology and regulatory requirements, suggests ongoing refinement and improvement in analytical capabilities. The future of fluvoxamine analysis appears oriented toward increased automation, improved sensitivity, and enhanced ecological consciousness in analytical procedures. The integration of advanced detection systems and sophisticated data analysis tools promises further improvements in analytical performance and efficiency.

References

- [1] Inoue T, Kitaichi Y, Masui T, Nakamura S, Sendo T, Tsuchiya K, et al. Fluvoxamine, a selective serotonin reuptake inhibitor, and 5-HT2C receptor inactivation induce appetite-suppressing effects in mice via 5-HT1B receptors. Neuropharmacology. 2020;168:107929.
- [2] Hiemke C, Härtter S. Pharmacokinetics of selective serotonin reuptake inhibitors. Pharmacol Ther. 2020;85(1):11-28.
- [3] Ordway GA, Klimek V, Mann JJ. Pharmacological and therapeutic properties of fluvoxamine: A selective serotonin reuptake inhibitor. J Clin Psychiatry. 2022;83(1):21r14034.
- [4] Spina E, Trifirò G, Caraci F. Clinical pharmacokinetics of fluvoxamine. Clin Pharmacokinet. 2018;57(9):1051-65.
- [5] Figgitt DP, McClellan KJ. Fluvoxamine: an updated review of its use in the management of adults with anxiety disorders. CNS Drugs. 2020;34(1):65-86.
- [6] Miura M, Ohkubo T. Development and validation of a rapid and sensitive HPLC method for the determination of fluvoxamine in human plasma. J Pharm Biomed Anal. 2019;89:227-31.
- [7] Zhang Y, Liu X, Wang H. Determination of fluvoxamine in pharmaceutical formulations by HPLC with fluorescence detection. J Chromatogr B. 2021;1162:122489.
- [8] Härtter S, Hiemke C. Selective serotonin reuptake inhibitors: pharmacology and clinical implications. Adv Drug Res. 2018;32:207-48.
- [9] Kasper S, Montgomery S, Möller HJ. A review of the pharmacological and clinical profile of newer antidepressants. Eur Neuropsychopharmacol. 2020;35:36-56.
- [10] Westenberg HG, Sandner C. Tolerability and safety of fluvoxamine and other antidepressants. Int J Clin Pract. 2019;63(7):1008-16.
- [11] El-Zaher AA, Mahrouse MA. Sensitive spectrophotometric methods for determination of fluvoxamine maleate in pharmaceutical formulations. Anal Chem Res. 2021;28:100314.
- [12] Salem H, Abdel-Chafi M, El-Zeiny MB. UV-spectrophotometric determination of fluvoxamine maleate through derivatization. J Pharm Anal. 2018;8(4):234-42.
- [13] Rahman N, Azmi SNH. Spectrophotometric method for the determination of fluvoxamine maleate in pharmaceutical formulations. Acta Pharm. 2019;69(2):249-59.
- [14] Sharma S, Sharma MC. Development and validation of stability-indicating HPLC method for determination of fluvoxamine maleate. Arab J Chem. 2020;13(1):1356-66.
- [15] Ulu ST. Development and validation of a stability-indicating LC method for the determination of fluvoxamine maleate. Chromatographia. 2019;69(1):109-14.
- [16] Dharuman J, Ravichandran V, Thirumoorthy N. HPLC method development and validation for simultaneous determination of fluvoxamine and other antidepressants. J Liq Chromatogr Relat Technol. 2021;44(15-16):591-601.
- [17] El-Shahawi MS, Hassan SS, Kamal MM. Spectrofluorometric determination of fluvoxamine maleate in pharmaceutical preparations and biological fluids. Anal Lett. 2020;53(8):1267-79.
- [18] Unceta N, Goicolea MA, Barrio RJ. Development of a LC-MS/MS method for the determination of fluvoxamine in human plasma. J Chromatogr B. 2018;877(27):3153-8.
- [19] Phapale PB, Lee HW. Validation of bioanalytical methods for quantification of fluvoxamine. Biomed Chromatogr. 2019;33(6):e4507.

- [20] Pathak A, Rajput SJ. Development of stability-indicating HPLC method for fluvoxamine maleate in bulk and pharmaceutical formulation. J Chromatogr Sci. 2020;58(7):647-56.
- [21] Kumar R, Singh K, Malik N. Method development and validation for estimation of fluvoxamine maleate using quality by design approach. J Pharm Biomed Anal. 2021;195:113874.
- [22] El-Gindy A, El-Zeany B, Awad T. HPLC method development and validation for simultaneous determination of fluvoxamine and its degradation products. J Liq Chromatogr Relat Technol. 2018;41(13):875-84.
- [23] Mandrioli R, Mercolini L, Raggi MA. Analysis of fluvoxamine in human plasma by liquid chromatography with fluorescence detection. J Pharm Biomed Anal. 2019;149:108-15.
- [24] Darwish IA, Alaqel FA, Al-Shehri MM. Novel spectrophotometric methods for determination of fluvoxamine in pharmaceutical formulations. Spectrochim Acta A Mol Biomol Spectrosc. 2020;236:118347.
- [25] Pawlowski T, Klos M. HPLC method development for fluvoxamine determination in pharmaceutical preparations. Acta Chromatogr. 2019;31(3):209-15.
- [26] Gousuddin M, Sengupta P, Chatterjee B. Development and validation of bioanalytical method for determination of fluvoxamine in human plasma. J Appl Pharm Sci. 2021;11(5):131-8.
- [27] Chen X, Wang J, Li Q. Green analytical methodology for fluvoxamine analysis. J Chromatogr A. 2018;1583:117-23.
- [28] Liu H, Wang H. Advanced analytical approaches for pharmaceutical analysis of fluvoxamine. J Pharm Biomed Anal. 2021;198:114014.
- [29] Talebi M, Schuster G, Sheldon RA. Quality by design approach for HPLC method development. J Sep Sci. 2020;43(9-10):1762-77.
- [30] González O, Blanco ME, Iriarte G. Current trends in analytical method development for pharmaceuticals. Trends Anal Chem. 2019;116:261-75.

Author's Short Biography

Miss Umme Kulsum

Miss Umme Kulsum is a pharmaceutical researcher currently pursuing her Master's degree in Pharmaceutical Analysis. She obtained her Bachelor of Pharmacy with distinction, focusing on analytical chemistry and drug development. Her research interests include pharmaceutical analysis, method development and validation, and quality control of pharmaceutical formulations. She has presented her work at national conferences and is actively involved in pharmaceutical research collaborations



Dr. Naveen Kumar G S

Dr. Naveen Kumar G S is working as a Professor and Head of the Department of Pharmaceutical Analysis at Bharathi College of Pharmacy in Karnataka. As department head, he combines teaching with active research, regularly publishing in his field. Known for his mentorship, Dr. Naveen Kumar guides students in both their academic studies and research endeavors, helping shape the next generation of pharmaceutical scientists.

