

## REVIEW ARTICLE

# Ultra-Performance Liquid Chromatography (UPLC) in Analytical Method Development and Validation Following *In-vitro* Studies



Ismail Y\*<sup>1</sup>, Jayadev Seelam<sup>2</sup>

<sup>1</sup>Associate Professor, Research Scholar, Crescent School of Pharmacy, B.S. Abdur Rahman Crescent Institute of Science and Technology, Vandalur, Chennai, Tamil Nadu, India

<sup>2</sup>Research Scholar, Crescent School of Pharmacy, B.S. Abdur Rahman Crescent Institute of Science and Technology, Vandalur, Chennai, Tamil Nadu, India

Publication history: Received on 1<sup>st</sup> Feb 2025; Revised on 14<sup>th</sup> Feb 2025; Accepted on 15<sup>th</sup> Feb 2025

Article DOI: 10.69613/rwktfg11

**Abstract:** Ultra-performance liquid chromatography (UPLC) has emerged as a powerful analytical tool, offering superior resolution, sensitivity, and rapid analysis compared to conventional high-performance liquid chromatography. The integration of UPLC with advanced detection systems, particularly mass spectrometry, has transformed pharmaceutical research, biomarker discovery, and metabolomics studies. Recent innovations in column technology, including sub-2-micron particles and core-shell materials, have significantly enhanced separation efficiency and detection limits. UPLC plays a crucial role in drug metabolism studies, stability assessments, and bioactive compound quantification following *in vitro* experiments. The technique has proven invaluable in toxicology studies, enabling precise identification and quantification of metabolites in biological matrices. Method validation parameters for UPLC ensure reliable and reproducible results, encompassing specificity, linearity, accuracy, precision, and stability studies. Notable applications include rapid metabolite profiling, protein characterization in biopharmaceuticals, and biomarker identification in disease diagnosis. Despite challenges such as operational costs and complex sample preparation requirements, ongoing technological advancements in microfluidics, artificial intelligence integration, and green chromatography techniques continue to expand UPLC capabilities.

**Keywords:** Ultra-performance liquid chromatography; Bioanalytical method validation; Drug metabolism; Metabolomics; Column.

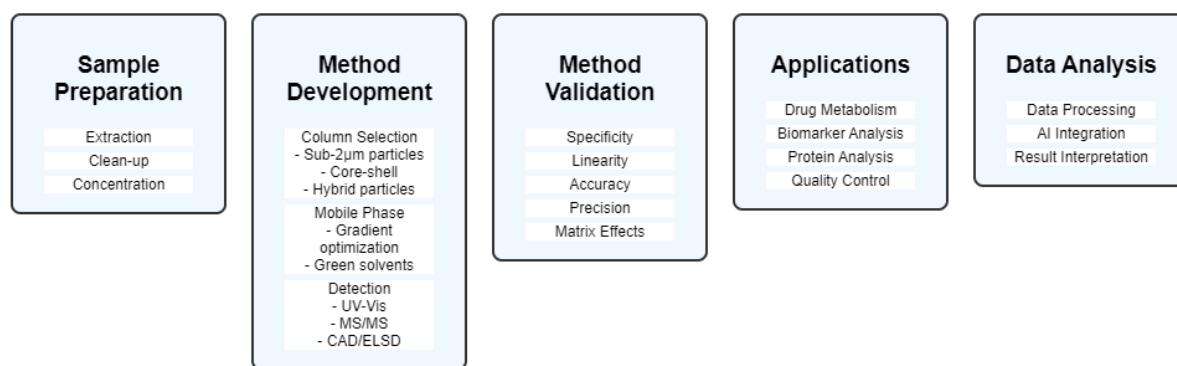
## 1. Introduction

The evolution of analytical techniques in pharmaceutical research has led to significant advancements in drug development and bioanalysis. Ultra-performance liquid chromatography (UPLC) represents a major technological leap in separation science, offering enhanced analytical capabilities compared to traditional high-performance liquid chromatography (HPLC) [1]. The transition from *in vitro* studies to comprehensive analytical evaluations necessitates robust methodologies that can generate reliable and reproducible results [2, 3].

UPLC employs innovative column technology and advanced instrumentation to achieve superior chromatographic performance. The use of sub-2-micron particle sizes and elevated operating pressures results in improved resolution, sensitivity, and dramatically reduced analysis times [4]. These attributes make UPLC particularly valuable in drug metabolism studies, stability testing, and bioactive compound quantification following *in vitro* experiments [5]. The integration of UPLC with mass spectrometry has further expanded its analytical capabilities, enabling simultaneous quantification and structural elucidation of complex biological compounds [6]. This technological synergy has proven especially beneficial in metabolomics research, biomarker discovery, and the characterization of biopharmaceuticals [7, 8].

UPLC serves as an essential tool in pharmaceutical analysis, particularly in drug development phases following *in vitro* studies. The technique enables rapid quantification of active pharmaceutical ingredients, related substances, and degradation products [9]. In dissolution studies, UPLC provides precise measurements of drug release profiles, contributing to the optimization of formulation parameters [10]. The enhanced resolution and sensitivity of UPLC facilitate the detection of trace-level impurities, ensuring compliance with regulatory requirements [11].

\* Corresponding author: Ismail Y and Jayadev Seelam

**Figure 1.** UPLC workflow

UPLC-MS systems excel in comprehensive metabolic profiling, offering detailed insights into cellular processes and metabolic pathways [12]. The technique can simultaneously analyze hundreds of metabolites in biological samples, making it invaluable for understanding drug-induced metabolic changes [13]. The high-throughput capability of UPLC enables rapid screening of metabolic fingerprints, accelerating biomarker discovery and validation [14]. In clinical research, UPLC has revolutionized the identification and quantification of disease-specific biomarkers [15]. The technique's ability to detect subtle changes in metabolite levels aids in early disease diagnosis and treatment monitoring. Advanced UPLC-MS methods have successfully identified novel biomarkers for various conditions, including cancer, cardiovascular diseases, and metabolic disorders [16].

UPLC plays a crucial role in toxicology studies by enabling the detection and quantification of toxic metabolites in biological systems [17]. The technique's high sensitivity allows for the identification of low-abundance toxic compounds and their metabolites in complex matrices [18]. Real-time monitoring of metabolic transformations helps elucidate toxicity mechanisms and assess potential drug-drug interactions [19].

**Table 1.** Common Applications of UPLC in Post-*In vitro* Studies

Application	Features	Typical Analysis Time	Detection Method
Drug Metabolism	Metabolite profiling	2-5 minutes	MS/MS
Protein Analysis	Peptide mapping	10-15 minutes	UV, MS
Biomarker Studies	Targeted quantification	3-8 minutes	MS
Stability Testing	Degradation products	5-10 minutes	UV, CAD
Impurity Profiling	Trace analysis	8-12 minutes	UV, MS

## 2. Methodological Advances in UPLC

### 2.1. Column Technology

Modern UPLC columns incorporate advanced particle technologies that significantly enhance separation efficiency. Sub-2-micron particles provide increased surface area and improved mass transfer characteristics [20]. Core-shell particles, featuring a solid core surrounded by a porous outer layer, minimize band broadening and enhance column efficiency [21]. The development of hybrid organic-inorganic particles has improved column stability under high-pressure conditions while maintaining separation performance across a broader pH range [22].

### 2.2. Mobile Phase Optimization

#### 2.2.1. Gradient Elution

Advanced gradient elution techniques optimize separation of complex mixtures from *in vitro* systems. Multi-step gradients enable resolution of compounds with similar physicochemical properties [23]. Temperature-assisted separation techniques further enhance selectivity and reduce analysis time [24].

#### 2.2.2. Green Chemistry

Environmental considerations have led to the development of sustainable mobile phase compositions. The incorporation of bio-based solvents and reduced organic solvent consumption aligns with green chemistry principles [25]. Novel aqueous mobile phase modifiers improve separation while minimizing environmental impact [26].

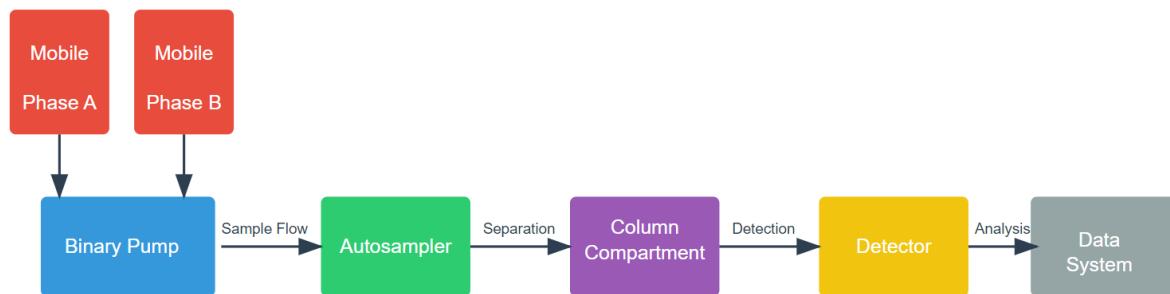


Figure 2. Components of HPLC

### 2.3. Detection Systems and Hyphenation

#### 2.3.1. Mass Spectrometry Integration

The coupling of UPLC with high-resolution mass spectrometry enables precise molecular characterization. Advanced MS interfaces facilitate efficient ionization and improved sensitivity [27]. Multi-stage MS capabilities allow detailed structural elucidation of unknown compounds and metabolites [28].

#### 2.3.2. Alternative Detection Methods

Novel detection systems include charged aerosol detection (CAD) and evaporative light scattering detection (ELSD), offering universal detection capabilities for non-chromophoric compounds [29]. Multi-detector arrangements provide complementary information for complex sample analysis [30].

Table 2. Comparison of Modern UPLC Column Technologies

Column Type	Particle Size	Pressure Limit	Advantages	Applications
Sub-2 $\mu$ m Fully Porous	1.7-1.8 $\mu$ m	15,000 psi	High efficiency	Small molecule analysis
Core-shell	2.6-2.7 $\mu$ m	9,000 psi	Reduced backpressure	Protein separations
Hybrid Particles	1.7-1.8 $\mu$ m	18,000 psi	pH stability	Multi-purpose analysis
Monolithic	N/A	8,000 psi	High throughput	Fast screening
Superficially Porous	2.0-2.7 $\mu$ m	12,000 psi	Sharp peaks	Complex mixtures

## 3. Bioanalytical Method Validation

### 3.1. Validation Parameters

#### 3.1.1. Specificity and Selectivity

Method development begins with establishing specificity for target analytes in complex biological matrices. Interference testing using blank matrix samples ensures reliable analyte identification [31]. The implementation of orthogonal separation mechanisms enhances selectivity for challenging separations [32].

#### 3.1.2. Linearity and Range

Calibration strategies employ appropriate concentration ranges reflecting expected analyte levels. Statistical evaluation of linearity includes assessment of correlation coefficients and residual plots [33]. The dynamic range encompasses at least three orders of magnitude for quantitative applications [34].

#### 3.1.3. Accuracy and Precision

Systematic evaluation of accuracy involves analysis of quality control samples at multiple concentration levels. Method precision assessment includes both intra-day and inter-day variability studies [35]. Robustness testing ensures method reliability under varying experimental conditions [36].

#### 3.1.4. Matrix Effects

Comprehensive evaluation of matrix effects involves post-column infusion studies and matrix factor determination [37]. The implementation of stable isotope-labeled internal standards compensates for matrix-induced response variations [38].

### 3.1.5. Stability Studies

Stability assessment encompasses multiple storage and handling conditions relevant to sample analysis. Long-term stability studies evaluate analyte stability under intended storage conditions [39]. Freeze-thaw stability and post-preparative stability ensure sample integrity throughout the analytical process [40].

**Table 3.** Validation Parameters and Acceptance Criteria

Parameter	Acceptance Criteria	Test Level	Number of Replicates
Specificity	No interference	N/A	Minimum 6
Linearity	$R^2 \geq 0.999$	6-8 levels	3 per level
Accuracy	98-102%	3 levels	6 per level
Precision (%RSD)	$\leq 2.0\%$	3 levels	6 per level
LOQ	$S/N \geq 10$	N/A	6

## 4. Practical Applications

### 4.1. Drug Metabolism

#### 4.1.1. Metabolite Identification

UPLC-MS/MS methodologies enable comprehensive metabolite profiling in complex biological matrices. Advanced data acquisition strategies, including neutral loss scanning and precursor ion monitoring, facilitate metabolite detection [41]. Real-time metabolite identification algorithms aid in structure elucidation and metabolic pathway mapping [42].

#### 4.1.2. Enzyme Kinetics

Detailed enzyme kinetic studies benefit from UPLC's rapid analysis capabilities. High-throughput screening of enzyme-substrate interactions provides valuable insights into drug biotransformation [43]. Time-course studies of metabolite formation enable accurate determination of kinetic parameters [44].

### 4.2. Biopharmaceutical Analysis

#### 4.2.1. Protein Characterization

UPLC methods excel in protein therapeutic characterization, including monoclonal antibodies and fusion proteins. Advanced peptide mapping techniques provide comprehensive sequence coverage and post-translational modification analysis [45]. Host cell protein analysis ensures product purity and safety [46].

#### 4.2.2. Quality Control

Routine quality control testing employs validated UPLC methods for product release. Size-variant analysis identifies aggregates and fragments in protein therapeutics [47]. Charge-variant analysis enables monitoring of product heterogeneity [48].

**Table 4.** Applications of UPLC in Biopharmaceutical Analysis

Analysis Type	Target Analytes	Method Parameters	Critical Attributes
Peptide Mapping	Digest fragments	Gradient elution	Sequence coverage
Glycan Analysis	N-linked glycans	HILIC mode	Glycoform distribution
Charge Variants	Protein isoforms	Ion exchange	Charge heterogeneity
Aggregates	Size variants	SEC mode	Aggregate content
PTM Analysis	Modified peptides	RP mode	Modification sites

### 4.3. Clinical Applications

#### 4.3.1. Therapeutic Drug Monitoring

UPLC enables rapid quantification of drugs and metabolites in biological fluids. Multi-analyte methods facilitate simultaneous monitoring of multiple therapeutic agents [49]. Advanced sample preparation techniques improve method sensitivity and specificity [50].

#### 4.3.2. Disease Biomarkers

Novel UPLC-based approaches aid in disease diagnosis and monitoring. Metabolomic profiling identifies disease-specific biomarker patterns [51]. Longitudinal studies track biomarker changes during disease progression and treatment [52].

---

## 5. Challenges and Limitations

### 5.1. Technical Aspects

The implementation of UPLC methods presents several technical challenges. High operating pressures require specialized instrumentation and maintenance protocols [53]. Sample matrix complexity necessitates careful method development and validation strategies [54].

### 5.2. Operational Aspects

#### 5.2.1. Cost Considerations

Initial investment in UPLC instrumentation represents a significant financial commitment. Ongoing operational costs include specialized columns and high-purity solvents [55]. Regular maintenance requirements contribute to overall system costs [56].

#### 5.2.2. Training

Successful UPLC implementation demands comprehensive operator training. Method development expertise requires substantial theoretical knowledge and practical experience [57]. Troubleshooting skills are essential for maintaining system performance [58].

---

## 6. Future Perspectives

### 6.1. Technological Advancements

#### 6.1.1. Artificial Intelligence

Machine learning algorithms enhance method development and optimization. Automated data processing improves metabolite identification accuracy [59]. Predictive maintenance systems optimize instrument performance [60].

#### 6.1.2. Miniaturization

Microfluidic UPLC systems offer reduced sample and solvent consumption. Novel column designs enable improved separation efficiency at microscale [61]. Integration with portable mass spectrometers advances field-based applications [62].

**Table 5.** Emerging Trends in UPLC

Technology	Innovation	Benefits
AI Integration	Automated method development	Reduced optimization time
Microfluidics	Chip-based separations	Minimal sample volume
Green Chemistry	Bio-based solvents	Environmental sustainability
Smart Diagnostics	IoT integration	Remote monitoring
Multi-dimensional	2D-UPLC	Enhanced resolution

#### 6.1.3. Sustainable Practices

Environmental considerations drive development of eco-friendly UPLC methods. Novel stationary phases reduce solvent consumption and waste generation [63]. Alternative mobile phase compositions minimize environmental impact [64].

---

## 7. Conclusion

Ultra-performance liquid chromatography has transformed analytical chemistry through its superior separation capabilities and versatile applications. The technique's continuous evolution, marked by advances in column technology, detection systems, and method validation strategies, has established its pivotal role in pharmaceutical and biomedical research. Integration with mass spectrometry and artificial intelligence promises enhanced analytical capabilities, while sustainable practices address environmental concerns.

## References

- [1] Swartz ME. UPLC™: An introduction and review. *J Liq Chromatogr Relat Technol.* 2005;28(7-8):1253-1263.
- [2] Nováková L, Matysová L, Solich P. Advantages of application of UPLC in pharmaceutical analysis. *Talanta.* 2006;68(3):908-918.
- [3] Zhang YJ, Wang XL, Xu B, et al. UPLC-MS/MS method for determination and pharmacokinetic study of davunetide in rat plasma. *J Chromatogr B.* 2014;967:183-188.
- [4] Wu T, Wang C, Wang X, et al. Comparison of UPLC and HPLC for analysis of system suitability tests. *J Pharm Anal.* 2015;5(4):223-230.
- [5] Fekete S, Schappler J, Veuthey JL, Guillarme D. Current and future trends in UHPLC. *TrAC Trends Anal Chem.* 2014;63:2-13.
- [6] Buszewski B, Noga S. Hydrophilic interaction liquid chromatography (HILIC)—a powerful separation technique. *Anal Bioanal Chem.* 2012;402(1):231-247.
- [7] Want EJ, Wilson ID, Gika H, et al. Global metabolic profiling procedures for urine using UPLC-MS. *Nat Protoc.* 2010;5(6):1005-1018.
- [8] Kumar A, Saini G, Nair A, Sharma R. UPLC: A preeminent technique in pharmaceutical analysis. *Acta Pol Pharm.* 2012;69(3):371-380.
- [9] Dong MW, Zhang K. Ultra-high-pressure liquid chromatography (UHPLC) in method development. *TrAC Trends Anal Chem.* 2014;63:21-30.
- [10] Wren SA, Tchelitcheff P. Use of ultra-performance liquid chromatography in pharmaceutical development. *J Chromatogr A.* 2006;1119(1-2):140-146.
- [11] Guillarme D, Nguyen DT, Rudaz S, Veuthey JL. Method transfer for fast liquid chromatography in pharmaceutical analysis. *Eur J Pharm Biopharm.* 2008;68(2):430-440.
- [12] Wilson ID, Nicholson JK, Castro-Perez J, et al. High resolution "ultra performance" liquid chromatography coupled to oa-TOF mass spectrometry as a tool for differential metabolic pathway profiling in functional genomic studies. *J Proteome Res.* 2005;4(2):591-598.
- [13] Plumb RS, Johnson KA, Rainville P, et al. UPLC/MSE; a new approach for generating molecular fragment information for biomarker structure elucidation. *Rapid Commun Mass Spectrom.* 2006;20(13):1989-1994.
- [14] Lei Z, Huhman DV, Sumner LW. Mass spectrometry strategies in metabolomics. *J Biol Chem.* 2011;286(29):25435-25442.
- [15] Zhang A, Sun H, Wang P, et al. Recent and potential developments of biofluid analyses in metabolomics. *J Proteomics.* 2012;75(4):1079-1088.
- [16] Theodoridis GA, Gika HG, Want EJ, Wilson ID. Liquid chromatography-mass spectrometry based global metabolite profiling: a review. *Anal Chim Acta.* 2012;711:7-16.
- [17] Ramirez T, Daneshian M, Kamp H, et al. Metabolomics in toxicology and preclinical research. *ALTEX.* 2013;30(2):209-225.
- [18] Dunn WB, Wilson ID, Nicholls AW, Broadhurst D. The importance of experimental design and QC samples in large-scale and MS-driven untargeted metabolomic studies of humans. *Bioanalysis.* 2012;4(18):2249-2264.
- [19] Churchwell MI, Twaddle NC, Meeker LR, Doerge DR. Improving LC-MS sensitivity through increases in chromatographic performance: Comparisons of UPLC-ES/MS/MS to HPLC-ES/MS/MS. *J Chromatogr B.* 2005;825(2):134-143.
- [20] MacNair JE, Lewis KC, Jorgenson JW. Ultrahigh-pressure reversed-phase liquid chromatography in packed capillary columns. *Anal Chem.* 1997;69(6):983-989.
- [21] Gritti F, Guiochon G. The van Deemter equation: assumptions, limits, and adjustment to modern high performance liquid chromatography. *J Chromatogr A.* 2013;1302:1-13.
- [22] Neue UD, O'Gara JE, Méndez A. Selectivity in reversed-phase separations: Influence of the stationary phase. *J Chromatogr A.* 2006;1127(1-2):161-174.
- [23] Snyder LR, Dolan JW. High-performance gradient elution: the practical application of the linear-solvent-strength model. John Wiley & Sons; 2007.
- [24] Carr PW, Stoll DR, Wang X. Perspectives on recent advances in the speed of high-performance liquid chromatography. *Anal Chem.* 2011;83(6):1890-1900.

[25] Gaber Y, Törnvall U, Kumar MA, et al. HPLC-ESI-MS/MS characterization of novel peptide derivatives from environmentally sustainable microwave-assisted synthesis. *Green Chem.* 2011;13(8):2021-2025.

[26] Welch CJ, Wu N, Biba M, et al. Greening analytical chromatography. *TrAC Trends Anal Chem.* 2010;29(7):667-680.

[27] Kaufmann A. Strategy for the elucidation of unknown pharmaceutical drug metabolism by electrospray ionisation liquid chromatography/tandem mass spectrometry. *J Chromatogr A.* 2009;1216(9):1474-1487.

[28] Holčapek M, Jirásko R, Lísá M. Recent developments in liquid chromatography–mass spectrometry and related techniques. *J Chromatogr A.* 2012;1259:3-15.

[29] Vervoort N, Daemen D, Török G. Performance evaluation of evaporative light scattering detection and charged aerosol detection in reversed phase liquid chromatography. *J Chromatogr A.* 2008;1189(1-2):92-100.

[30] Lesellier E, West C. The many faces of packed column supercritical fluid chromatography – A critical review. *J Chromatogr A.* 2015;1382:2-46.

[31] Rozet E, Marini RD, Ziemons E, et al. Advances in validation, risk and uncertainty assessment of bioanalytical methods. *J Pharm Biomed Anal.* 2011;55(4):848-858.

[32] Guillarme D, Ruta J, Rudaz S, Veuthey JL. New trends in fast and high-resolution liquid chromatography: a critical comparison of existing approaches. *Anal Bioanal Chem.* 2010;397(3):1069-1082.

[33] González AG, Herrador MÁ. A practical guide to analytical method validation, including measurement uncertainty and accuracy profiles. *TrAC Trends Anal Chem.* 2007;26(3):227-238.

[34] Matuszewski BK, Constanzer ML, Chavez-Eng CM. Strategies for the assessment of matrix effect in quantitative bioanalytical methods based on HPLC–MS/MS. *Anal Chem.* 2003;75(13):3019-3030.

[35] Shah VP, Midha KK, Findlay JW, et al. Bioanalytical method validation—a revisit with a decade of progress. *Pharm Res.* 2000;17(12):1551-1557.

[36] Peters FT, Drummer OH, Musshoff F. Validation of new methods. *Forensic Sci Int.* 2007;165(2-3):216-224.

[37] Taylor PJ. Matrix effects: the Achilles heel of quantitative high-performance liquid chromatography–electrospray–tandem mass spectrometry. *Clin Biochem.* 2005;38(4):328-334.

[38] Stokvis E, Rosing H, Beijnen JH. Stable isotopically labeled internal standards in quantitative bioanalysis using liquid chromatography/mass spectrometry: necessity or not? *Rapid Commun Mass Spectrom.* 2005;19(3):401-407.

[39] Nowatzke W, Woolf E. Best practices during bioanalytical method validation for the characterization of assay reagents and the evaluation of analyte stability in assay standards, quality controls, and study samples. *AAPS J.* 2007;9(2):E117-E122.

[40] Mangam VT, Narla D, Konda RK, Sarella PN. Beyond the spectrum: Exploring unconventional applications of fourier transform infrared (FTIR) spectroscopy. *Asian Journal of Pharmaceutical Analysis.* 2024;14(2):86-94.

[41] Zhu M, Zhang H, Humphreys WG. Drug metabolite profiling and identification by high-resolution mass spectrometry. *J Biol Chem.* 2011;286(29):25419-25425.

[42] Zhang H, Yang Y. An algorithm for thorough background subtraction from high-resolution LC/MS data: application for detection of glutathione-trapped reactive metabolites. *J Mass Spectrom.* 2008;43(9):1181-1190.

[43] Trunzer M, Faller B, Zimmerlin A. Metabolic soft spot identification and compound optimization in early discovery phases using MetaSite and LC-MS/MS validation. *J Med Chem.* 2009;52(2):329-335.

[44] Brandon EFA, Raap CD, Meijerman I, et al. An update on in vitro test methods in human hepatic drug biotransformation research: pros and cons. *Toxicol Appl Pharmacol.* 2003;189(3):233-246.

[45] Beck A, Wagner-Rousset E, Ayoub D, et al. Characterization of therapeutic antibodies and related products. *Anal Chem.* 2013;85(2):715-736.

[46] Wang F, Richardson D, Shameem M. Host-cell protein measurement and control. *BioPharm Int.* 2015;28(6):32-38.

[47] Fekete S, Guillarme D, Sandra P, Sandra K. Chromatographic, electrophoretic, and mass spectrometric methods for the analytical characterization of protein biopharmaceuticals. *Anal Chem.* 2016;88(1):480-507.

[48] Zhang T, Yang X, Xu W, et al. Advances in chromatographic analysis of therapeutic proteins. *TrAC Trends Anal Chem.* 2016;81:151-164.

[49] Patel DS, Sharma N, Patel MC, et al. Development and validation of a selective and sensitive LC–MS/MS method for determination of cycloserine in human plasma: application to bioequivalence study. *J Chromatogr B.* 2011;879(23):2265-2273.

- [50] Kole PL, Venkatesh G, Kotecha J, Sheshala R. Recent advances in sample preparation techniques for effective bioanalytical methods. *Biomed Chromatogr*. 2011;25(1-2):199-217.
- [51] Psychogios N, Hau DD, Peng J, et al. The human serum metabolome. *PLoS One*. 2011;6(2):e16957.
- [52] Dunn WB, Broadhurst DI, Atherton HJ, et al. Systems level studies of mammalian metabolomes: the roles of mass spectrometry and nuclear magnetic resonance spectroscopy. *Chem Soc Rev*. 2011;40(1):387-426.
- [53] Arji SR, Eranki SS, Pecchetty S, Sarella PN. Unconventional stationary phases: Nanomaterials, nanoparticles and the future of liquid chromatography. *World Journal of Advanced Research and Reviews*. 2023;18(2):492-501.
- [54] Nováková L, Vlčková H. A review of current trends and advances in modern bio-analytical methods: chromatography and sample preparation. *Anal Chim Acta*. 2009;656(1-2):8-35.
- [55] Walter TH, Andrews RW. Recent innovations in UHPLC columns and instrumentation. *TrAC Trends Anal Chem*. 2014;63:14-20.
- [56] Guillarme D, Ruta J, Rudaz S, Veuthey JL. New trends in fast and high-resolution liquid chromatography: a critical comparison of existing approaches. *Anal Bioanal Chem*. 2010;397(3):1069-1082.
- [57] Kromidas S. *HPLC made to measure: a practical handbook for optimization*. John Wiley & Sons; 2008.
- [58] Snyder LR, Kirkland JJ, Dolan JW. *Introduction to modern liquid chromatography*. John Wiley & Sons; 2011.
- [59] Beccaria M, Bobak C, Maitshotlo B, et al. Analytical characterization of human exhaled breath condensate: A review. *J Breath Res*. 2019;13(3):034001.
- [60] Parasuraman S, Walker S, Loudon BL, et al. Assessment of pulmonary artery pressure by echocardiography—A comprehensive review. *Int J Cardiol Heart Vasc*. 2016;12:45-51.
- [61] Gama MR, Collins CH, Bottoli CB. Nano-liquid chromatography in pharmaceutical and biomedical research. *J Chromatogr Sci*. 2013;51(7):694-703.
- [62] Gehrke CW, Wixom RL, Bayer E. *Chromatography: a century of discovery 1900-2000*. Elsevier; 2001.
- [63] Welch CJ, Brkovic T, Schafer W, Gong X. Performance to burn? Re-evaluating the choice of acetonitrile as the platform solvent for analytical HPLC. *Green Chem*. 2009;11(8):1232-1238.
- [64] Hartonen K, Riekkola ML. Liquid chromatography at elevated temperatures with pure water as the mobile phase. *TrAC Trends Anal Chem*. 2008;27(1):1-14.