REVIEW ARTICLE

A Review of Recent Advances in the Classification, Pathogenesis, Diagnosis, and Treatment of Idiopathic Inflammatory Myopathies



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Abstract: Idiopathic inflammatory myopathies (IIM) represent a heterogeneous group of rare autoimmune disorders affecting skeletal muscles. These conditions are traditionally classified into three main subtypes: polymyositis (PM), dermatomyositis (DM), and inclusion body myositis (IBM). Recent advances in understanding their pathogenesis have led to the identification of new subtypes and associated autoantibodies, revolutionizing disease classification and treatment approaches. The association between genetic susceptibility, particularly HLA associations, and environmental triggers contributes significantly to disease development. Myositis-specific autoantibodies have emerged as crucial tools for diagnosis, prognosis, and therapeutic decision-making. Advanced diagnostic techniques, including muscle biopsy, imaging, and serological testing, offer enhanced clinical utility in disease identification and monitoring. Contemporary treatment strategies encompass conventional immunosuppressive therapy and novel targeted biological agents. Emerging therapies targeting specific pathways, including B-cell depletion, interferon signaling, and complement inhibition, show promising results in clinical studies. Early diagnosis and personalized treatment approaches based on disease subtypes and biomarker profiles have become fundamental principles in managing these disorders. The integration of novel diagnostic tools and targeted therapies has significantly improved the potential for better outcomes in patients with IIM, though challenges in treatment optimization remain.

Keywords: Idiopathic inflammatory myopathies; Polymyositis; Dermatomyositis; Creatine kinase; Autoimmune muscle disease.

1. Introduction

Idiopathic inflammatory myopathies (IIM) encompass a diverse group of rare autoimmune disorders primarily affecting skeletal muscles. For decades, these conditions have been categorized into three main subtypes based on distinct histological and clinical features: polymyositis (PM), dermatomyositis (DM), and inclusion body myositis (IBM) [1]. The limited therapeutic success and disease rarity often result in patients experiencing reduced mobility, progressive muscle weakness, and diminished quality of life, highlighting an unmet need for innovative treatment strategies [2]. Molecular pathway analysis has revealed distinct mechanisms in the early and chronic phases of myositis, crucial for developing targeted therapies. The three IIM subgroups exhibit varying clinical manifestations and therapeutic responses, suggesting predominant but distinct molecular pathways in each condition [3]. However, the traditional subgroup classification has limitations, as evidenced by overlapping muscle biopsy characteristics between PM and IBM patients, as well as between DM and PM cases [4]. A significant advancement in the past decade has been the recognition of IIM cases presenting predominantly with extramuscular manifestations, such as skin rashes, arthritis, or interstitial lung disease (ILD), even in the absence of muscle symptoms, as seen in amyopathic dermatomyositis [5]. The development of the EULAR/ACR classification criteria in 2017 marked a crucial milestone, providing a probability-based scoring system for adult and juvenile-onset IIM classification [6].

The identification of novel autoantibodies has substantially enhanced our understanding of IIM spectrum disorders. These autoantibodies fall into two categories: myositis-specific autoantibodies (MSA), predominantly found in IIM patients, and myositis-associated antibodies (MAA), which also occur in other autoimmune conditions like Sjögren's syndrome and systemic lupus erythematosus [7]. While immunoprecipitation has been valuable in identifying novel antigen specificities, its clinical application remains limited due to cost and technical demands. Recent developments in ELISA and line blot assays have made serum testing

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more accessible in clinical settings, though validation of these autoantibody assays requires international collaboration [8]. The complexity of IIM pathogenesis involves multiple factors, including genetic predisposition, environmental triggers, and immunological mechanisms. Recent research has highlighted the role of complement activation, interferon signaling, and B-cell-mediated responses in disease development [9].

Inflammatory Myopathies Classification & Clinical Presentations

Dermatomyositis

- Skin rash (Gottron's papules)
- Proximal muscle weakness
- Heliotrope rash
- Calcinosis
- · Interstitial lung disease
- · Cancer association

Polymyositis

- Symmetric proximal weakness
- · No skin involvement
- · Adult onset
- Dysphagia
- · Elevated CK levels
- · T-cell mediated damage

Inclusion Body Myositis

- Distal and proximal weakness
- · Finger flexor weakness
- · Quadriceps weakness
- Asymmetric involvement
- Slow progressionPoor treatment response

Immune-Mediated Necrotizing Myopathy

- Severe weakness
- High CK levels
- · Anti-SRP antibodies
- · Anti-HMGCR antibodies
- · Statin association
- · Minimal inflammation

Figure 1. Overview of IIM classification and clinical presentations

2. Classification

2.1. Dermatomyositis (DM)

Dermatomyositis represents a distinct subset of IIM with a reported prevalence of 1.4 to 5.8 cases per 100,000 individuals in the United States. The condition demonstrates a female predominance and increased occurrence in older populations [10]. Among juvenile cases, girls account for a significant proportion of the estimated 3.2 million affected children in the United Kingdom [11].

The clinical presentation of DM is characterized by distinctive erythematous changes accompanied by symmetric proximal muscle weakness that develops over weeks to months. Cutaneous manifestations, which may precede or follow myopathy, include pathognomonic features such as:

- Heliotrope rash
- Periorbital edema
- Mechanic's hands
- Gottron's papules over extensor surfaces
- Subcutaneous calcifications

Clinically amyopathic dermatomyositis (CADM), a distinct variant comprising approximately 20% of DM cases, presents with characteristic cutaneous changes but minimal or absent myopathy. CADM carries a significant risk of interstitial lung disease (ILD), particularly in cases positive for anti-CADM-140 antibodies [12]. The pathogenesis of DM involves a complex immunological cascade. Initially, immune complexes bind to endothelial cells, triggering complement activation and membrane attack complex (MAC)-mediated cell lysis [13]. This process results in i. Endothelial cell necrosis, ii. Significant reduction in muscle capillary density and iii. Perifascicular atrophy due to reduced blood flow

Recent research has challenged this traditional understanding by proposing a type-I interferon-mediated pathway. The overexpression of type I interferon (α/β) genes in blood, muscle, and skin correlates with disease activity, with dendritic cells potentially serving as interferon sources [14].

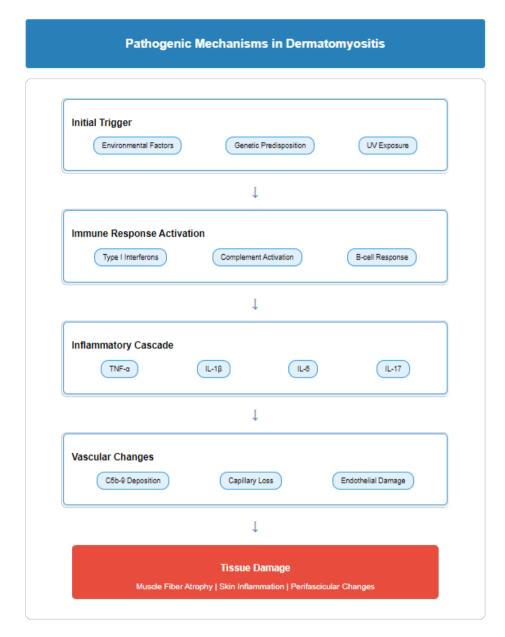


Figure 2. Pathogenic mechanisms in dermatomyositis

The inflammatory cascade in DM tissues involves increased expression of:

- Pro-inflammatory mediators (TGF-β, MHC-I, IL-1β)
- Chemokines (CCL-3, CCL-4)
- Adhesion molecules (VCAM-1, ICAM-1)

These factors facilitate immune cell migration and accumulation in affected tissues, particularly plasmacytoid dendritic cells, macrophages, B-cells, and T-cells in perivascular and perimysial regions [15].

2.2. Polymyositis (PM)

2.2.1. Clinical Presentation

Polymyositis manifests with marked elevation in creatine kinase (CK) levels and proximal muscle weakness, which may develop subacutely. The condition predominantly affects the shoulder and pelvic girdle musculature, with occasional involvement of neck flexors and extensors. Epidemiological data from the United States indicates an overall incidence rate of 9.7 per 100,000 persons, adjusting to 3.8 when accounting for age and gender variations [16].

2.2.2. Diagnostic Considerations

The diagnosis of PM requires careful consideration, as several studies suggest potential overdiagnosis when muscle biopsy confirmation is not employed. Histologically, cellular infiltrates comprising macrophages and cytotoxic CD8+ T lymphocytes are characteristic. A distinguishing feature from DM is the ability of these cells to surround and invade non-necrotic muscle fibers [17].

2.2.3. Immunopathological Mechanisms

The immunological environment in PM is characterized by the activation of immune cells within specific skeletal muscle regions. This activation triggers the production of various inflammatory mediators, including cytokines (IFN-γ, IL-6, IL-1β, TNF-α, TGF-β) and chemokines (IL-8, CCL-2, CCL-3, CCL-4, CCL-5, CXCL-9, CXCL-10). These factors create a pro-inflammatory milieu that promotes immune cell recruitment and local inflammation [18].

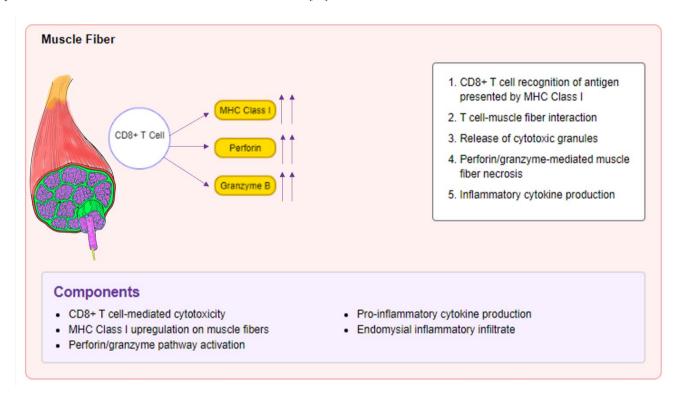


Figure 3. Immunopathological mechanisms in Polymyositis

2.3. Necrotizing Myopathy (NM)

2.3.1. Clinical Features

Necrotizing myopathy presents with progressive, symmetric weakness of proximal limb muscles. The condition, sometimes referred to as immune-mediated necrotizing myopathy, may include myalgia in up to 80% of cases. Severe presentations can involve dysphagia and dysarthria [19].

2.3.2. Pathological Subtypes

NM encompasses various disorders, including autoimmune inflammatory conditions, paraneoplastic diseases, and toxin-induced myopathies. Approximately 4-6% of myositis patients develop specific autoantibodies targeting either signal recognition particle or 3-hydroxy-3-methylglutaryl-coenzyme A reductase (HMGCR) [20].

2.3.3. Histopathological Features

Muscle biopsy reveals scattered necrotic muscle fibers as the predominant finding. Inflammatory cells, primarily macrophages with sparse CD4+ and CD8+ T-cells, may be dispersed throughout necrotic areas. The expression of MHC-I appears non-specific to either necrotic or regenerating fibers [21].

Table 1. Comparison of clinical and histological features across different types of myositis

Feature	Dermatomyositis	Polymyositis	Immune-Mediated Necrotizing Myopathy	Inclusion Body Myositis
Age of onset	Any age	Adult	Adult	>50 years
Gender ratio (F:M)	2:1	2:1	2:1	1:3
Clinical features	Proximal muscle weakness; Skin rash; Heliotrope rash; Gottron's papules; Dysphagia	Proximal muscle weakness; Dysphagia; No skin rash	Severe proximal weakness; Rapid progression; High CK levels	Slowly progressive; Distal > proximal weakness; Quadriceps and finger flexors; Falls
Muscle biopsy findings	Perifascicular atrophy; Perivascular inflammation; B and CD4+ T cells	Endomysial inflammation; CD8+ T cells; MHC-I upregulation	Myofiber necrosis; Minimal inflammation; MAC deposition	Rimmed vacuoles; Protein accumulation; Endomysial inflammation
Associated conditions	Cancer; Interstitial lung disease; Anti-synthetase syndrome	Anti-synthetase syndrome; Connective tissue diseases	Anti-HMGCR; Anti- SRP; Statin exposure	Usually isolated; Rare associated conditions
Treatment response	Good	Moderate	Variable	Poor
Key autoantibodies	Anti-Mi2; Anti-TIF1γ; Anti- NXP2; Anti-SAE; Anti- MDA5	Anti-Jo1; Other anti- synthetases	Anti-HMGCR; Anti- SRP	Anti-cN1A

CK: Creatine Kinase; MAC: Membrane Attack Complex; HMGCR: 3-Hydroxy-3-Methylglutaryl-Coenzyme A Reductase; SRP: Signal Recognition Particle; MHC: Major Histocompatibility Complex

2.4. Inclusion Body Myositis (IBM)

2.4.1. Epidemiology and Clinical Course

Inclusion body myositis emerges as the most prevalent acquired myopathy in individuals over fifty years of age. Australian epidemiological studies report an overall population frequency of 9.3 per million, escalating to 51.3 per million in the over-50 age group. Unlike PM and DM, IBM demonstrates a distinct gender distribution pattern [22].

2.4.2. Disease Progression

The condition typically presents with gradual, asymmetric muscle weakness developing over years. Characteristic features include selective involvement of knee extensors and finger flexors. Dysphagia affects 65-80% of IBM patients, ranging from mild to moderate severity, and may precede appendicular weakness. Disease progression typically impacts mobility, with patients requiring assistive devices approximately 14-16 years after symptom onset. Complete wheelchair dependency usually occurs after 24 years [23].

2.4.3. Diagnostic Criteria and Histopathology

The European Neuromuscular Centre (ENMC) has established diagnostic criteria based on histological findings and specific clinical parameters. Muscle biopsy reveals a combination of inflammatory processes and degenerative changes. The inflammatory component mirrors PM, with cytotoxic CD8+ T-cells and macrophages infiltrating non-necrotic fibers. Distinctive degenerative features include ringed vacuoles and intracellular β-amyloid deposits, detectable through Congo red or thioflavin-S staining [24].

2.4.4. Molecular Pathophysiology

The complex pathophysiology of IBM involves multiple protein accumulations, including gelsolin, clusterin, α -synuclein, apolipoprotein, γ -tubulin, P-tau, and presenilin 1. Cellular stress appears crucial in disease development, evidenced by the colocalization of APP/ β -amyloid and α B-crystallin. Pro-inflammatory conditions may promote fiber death through increased inducible nitric oxide synthase activity. Abnormal protein accumulation has been linked to macrophage processing, particularly under pro-inflammatory conditions [25].

3. Genetic Risk Factors

3.1.1. HLA Associations

Genome-wide single-nucleotide polymorphism (SNP) studies in European adults and juveniles with DM or PM have revealed strong disease associations within the MHC region on chromosome 6. The development of the Immunochip has enabled more precise analysis of HLA alleles and amino acids from SNP data [26].

The Myositis Genetics Consortium conducted the largest genetic study to date, involving 2,566 individuals from 14 countries. Results demonstrated strong associations between polymyositis and alleles of the 8.1 ancestral haplotype (HLA-DRB103:01), while dermatomyositis showed strong links to HLA-B08:01. Conditional analysis revealed independent disease associations within this haplotype [27].

Genetic risk factors demonstrate ethnic variability. In Chinese populations, dermatomyositis associates with HLA-DQA101:04 and HLA-DRB107, while Japanese populations show IIM associations with HLA-DRB1*08:03. These variations highlight the importance of considering ethnic background in genetic risk assessment [28]

4. Immunological Mechanisms

4.1.1. Innate Immune Response

Type I Interferon Signature: The role of type I interferons emerges as central to myositis pathogenesis. Muscle tissue analysis reveals heightened expression of type I interferon-inducible genes and proteins. Plasmacytoid dendritic cells, primary producers of type I interferons, accumulate in affected muscle tissue and skin lesions. These cells respond to immune complexes containing nucleic acids, triggering interferon production through toll-like receptor activation [29].

Complement System Involvement: Research demonstrates significant complement pathway activation in dermatomyositis. The membrane attack complex deposits on endomysial capillaries precede other inflammatory changes. This process leads to capillary destruction, perifascicular atrophy, and subsequent tissue damage. The role of complement extends beyond vascular injury, potentially influencing broader inflammatory processes [30].

4.1.2. Adaptive Immune Response

T Cell-Mediated Mechanisms: In polymyositis and inclusion body myositis, CD8+ T cells exhibit clonal expansion and perforindependent cytotoxicity against muscle fibers. These cells recognize antigens presented by MHC class I molecules, abnormally expressed on muscle fibers. The persistent presence of these expanded T cell populations suggests an antigen-driven immune response [31].

B Cell Participation: Recent investigations highlight the crucial role of B cells beyond antibody production. These cells function as antigen-presenting cells and cytokine producers, contributing to disease pathogenesis. The success of B cell depletion therapy in some patients underscores their pathogenic significance [32].

5. Autoantibodies In Myositis

5.1. Diagnostic Value

Myositis-specific autoantibodies serve as valuable diagnostic markers, often correlating with distinct clinical phenotypes. These autoantibodies target various cellular components, including aminoacyl-tRNA synthetases, nuclear proteins, and cytoplasmic elements. Their presence helps identify specific disease subsets and guides therapeutic decisions [33].

5.2. Prognostic Implications

The presence of specific autoantibodies carries significant prognostic value. Anti-synthetase antibodies associate with interstitial lung disease, while anti-Mi-2 antibodies generally indicate better treatment response. Anti-SRP antibodies correlate with severe necrotizing myopathy, often requiring aggressive immunotherapy [34].

5.3. Novel Autoantibodies

Advanced immunological techniques have led to the identification of new autoantibodies. Anti-TIF1 γ antibodies show strong association with cancer-associated dermatomyositis. Anti-NXP2 antibodies correlate with increased calcinosis risk in juvenile dermatomyositis. These discoveries continue to refine disease classification and risk assessment [35]

6. Diagnosis And Assessment

6.1. Physical Examination

Comprehensive musculoskeletal assessment remains fundamental in myositis diagnosis. Manual muscle testing using the Medical Research Council scale provides standardized strength measurement across muscle groups. The physical examination must include careful evaluation of skin changes, particularly in suspected dermatomyositis cases. Assessment of functional capacity through timed tests offers objective measurement of disease impact [36].

6.2. Serum Markers

Creatine kinase elevation serves as a primary indicator of muscle damage, though levels may not correlate directly with disease severity. Additional muscle enzymes, including aldolase, aspartate aminotransferase, and lactate dehydrogenase, provide supplementary information. Regular monitoring of these parameters helps track disease activity and treatment response [37].

6.3. Magnetic Resonance Imaging

MRI emerges as an invaluable tool in myositis diagnosis and monitoring. T2-weighted and STIR sequences reveal muscle edema, while T1-weighted images demonstrate fatty replacement and atrophy. Pattern recognition of muscle involvement aids in distinguishing between different myositis subtypes. Contemporary protocols incorporate whole-body MRI for comprehensive assessment [38].

6.4. Muscle Biopsy

Muscle biopsy remains the gold standard for definitive diagnosis. Site selection typically favors moderately affected muscles, avoiding severely weak or recently examined areas. Immunohistochemical staining enables detailed characterization of inflammatory infiltrates and assessment of muscle fiber protein expression patterns [39].

7. Treatment

7.1. Glucocorticoids

Initial treatment typically involves high-dose glucocorticoids, with subsequent dose adjustment based on clinical response. The starting dose usually ranges between 0.5-1 mg/kg/day of prednisone equivalent. Response assessment occurs at regular intervals, with dose reduction initiated after clinical improvement. Long-term steroid use necessitates careful monitoring for complications [40].

7.2. Immunosuppressive Agents

Second-line agents, including methotrexate, azathioprine, and mycophenolate mofetil, serve as steroid-sparing alternatives. Selection depends on individual patient factors, potential side effects, and specific disease manifestations. Regular monitoring of blood parameters ensures early detection of potential complications [41].

7.3. Novel Therapeutic Approaches

Rituximab demonstrates efficacy in refractory cases, particularly in antisynthetase syndrome and dermatomyositis. Clinical trials exploring other biological agents target specific pathogenic pathways, including type I interferon signaling and complement activation. These approaches represent a shift toward personalized medicine based on individual disease mechanisms [42].

7.4. Physical Rehabilitation

Structured exercise programs play a crucial role in maintaining muscle function and preventing contractures. Contemporary approaches combine resistance training with aerobic exercise, tailored to individual patient capabilities. Regular assessment ensures appropriate progression and prevents overexertion [43].

8. Prognosis

The outcome of inflammatory myopathies varies significantly among subtypes. Dermatomyositis typically demonstrates a biphasic pattern, with early response to therapy followed by potential relapses. Polymyositis often exhibits a more chronic course requiring sustained immunosuppression. Inclusion body myositis shows progressive deterioration despite therapeutic intervention [44].

8.1. Clinical Predictors

Several factors influence disease outcomes. Early diagnosis and treatment initiation correlate with improved prognosis. The presence of extramuscular manifestations, particularly interstitial lung disease, significantly impacts survival rates. Advanced age at onset and delayed diagnosis associate with poorer outcomes. Specific autoantibody profiles provide valuable prognostic information [45].

8.2. Quality of Life Impact

Long-term follow-up studies reveal substantial impact on daily activities and employment status. The Health Assessment Questionnaire Disability Index (HAQ-DI) demonstrates persistent functional limitation in many patients. Regular assessment of quality of life metrics guides therapeutic decision-making and rehabilitation strategies [46].

9. Conclusion

Inflammatory myopathies represent a complex spectrum of disorders requiring sophisticated diagnostic approaches and individualized therapeutic strategies. Recent advances in molecular understanding have revolutionized disease classification and treatment selection, particularly through the identification of specific autoantibodies and their clinical associations. The integration of conventional therapies with novel biological agents offers improved outcomes for many patients, though significant challenges remain in treating certain subtypes, notably inclusion body myositis. International collaborative efforts continue to enhance our understanding of disease mechanisms and therapeutic responses through standardized assessment protocols and large-scale patient registries. The future of myositis management lies in personalized medicine approaches, combining molecular profiling with targeted therapeutic interventions to optimize patient outcomes.

References

- [1] Selva-O'Callaghan A, Pinal-Fernandez I, Trallero-Araguás E, et al. Classification and management of adult inflammatory myopathies. Lancet Neurol. 2018;17(9):816-828.
- [2] Mariampillai K, Granger B, Amelin D, et al. Development of a new classification system for idiopathic inflammatory myopathies based on clinical manifestations and myositis-specific autoantibodies. JAMA Neurol. 2018;75(12):1528-1537.
- [3] Lundberg IE, Tjärnlund A, Bottai M, et al. 2017 European League Against Rheumatism/American College of Rheumatology classification criteria for adult and juvenile idiopathic inflammatory myopathies. Ann Rheum Dis. 2017;76(12):1955-1964.
- [4] Schmidt J. Current classification and management of inflammatory myopathies. J Neuromuscul Dis. 2018;5(2):109-129.
- [5] Milone M. Diagnosis and management of immune-mediated myopathies. Mayo Clin Proc. 2017;92(5):826-837.
- [6] Day J, Patel S, Limaye V. The role of magnetic resonance imaging techniques in evaluation and management of the idiopathic inflammatory myopathies. Semin Arthritis Rheum. 2017;46(5):642-649.
- [7] Miller FW, Lamb JA, Schmidt J, et al. Risk factors and disease mechanisms in myositis. Nat Rev Rheumatol. 2018;14(5):255-268.
- [8] Betteridge Z, McHugh N. Myositis-specific autoantibodies: an important tool to support diagnosis of myositis. J Intern Med. 2016;280(1):8-23.
- [9] Mammen AL. Autoimmune myopathies: autoantibodies, phenotypes and pathogenesis. Nat Rev Neurol. 2011;7(6):343-354.
- [10] Dalakas MC. Pathophysiology of inflammatory and autoimmune myopathies. Presse Med. 2011;40(4 Pt 2):e237-247.
- [11] Verschuuren JJ, Badrising UA, Van Engelen BG, et al. Inclusion body myositis. In: Handbook of Clinical Neurology. Vol 113. Elsevier; 2013:1311-1336.
- [12] Rider LG, Werth VP, Huber AM, et al. Measures of adult and juvenile dermatomyositis, polymyositis, and inclusion body myositis. Arthritis Care Res. 2011;63(S11):S118-S157.
- [13] Rothwell S, Cooper RG, Lundberg IE, et al. Dense genotyping of immune-related loci in idiopathic inflammatory myopathies confirms HLA alleles as the strongest genetic risk factor and suggests different genetic background for major clinical subgroups. Ann Rheum Dis. 2016;75(8):1558-1566.
- [14] Pinal-Fernandez I, Casal-Dominguez M, Mammen AL. Immune-mediated necrotizing myopathy. Curr Rheumatol Rep. 2018;20(4):21.
- [15] Selva-O'Callaghan A, Trallero-Araguás E, Grau-Junyent JM, et al. Cancer-associated myositis: clinical features and novel autoantibodies. Curr Opin Rheumatol. 2016;28(6):592-600.

- [16] Dobloug C, Garen T, Bitter H, et al. Prevalence and clinical characteristics of adult polymyositis and dermatomyositis. Ann Rheum Dis. 2015;74(8):1551-1556.
- [17] Dalakas MC. Inflammatory muscle diseases. N Engl J Med. 2015;372(18):1734-1747.
- [18] De Paepe B, Creus KK, De Bleecker JL. Role of cytokines and chemokines in idiopathic inflammatory myopathies. Curr Opin Rheumatol. 2009;21(6):610-616.
- [19] Allenbach Y, Mammen AL, Benveniste O, et al. 224th ENMC International Workshop: Clinico-sero-pathological classification of immune-mediated necrotizing myopathies. Neuromuscul Disord. 2018;28(1):87-99.
- [20] Watanabe Y, Uruha A, Suzuki S, et al. Clinical features and prognosis in anti-SRP and anti-HMGCR necrotising myopathy. J Neurol Neurosurg Psychiatry. 2016;87(10):1038-1044.
- [21] Allenbach Y, Benveniste O, Goebel HH, et al. Integrated classification of inflammatory myopathies. Neuropathol Appl Neurobiol. 2017;43(1):62-81.
- [22] Badrising UA, Maat-Schieman M, van Duinen SG, et al. Epidemiology of inclusion body myositis in the Netherlands. Neurology. 2000;55(9):1385-1387.
- [23] Rose MR, Group EIW. 188th ENMC International Workshop: Inclusion Body Myositis. Neuromuscul Disord. 2013;23(12):1044-1055.
- [24] Benveniste O, Stenzel W, Hilton-Jones D, et al. Amyloid deposits and inflammatory infiltrates in sporadic inclusion body myositis. Brain. 2015;138(Pt 5):1442-1454.
- [25] Greenberg SA. Inclusion body myositis: clinical features and pathogenesis. Nat Rev Rheumatol. 2019;15(5):257-272.
- [26] Miller FW, Chen W, O'Hanlon TP, et al. Genome-wide association study identifies HLA 8.1 ancestral haplotype alleles as major genetic risk factors for myositis phenotypes. Genes Immun. 2015;16(7):470-480.
- [27] Rothwell S, Cooper RG, Lundberg IE, et al. Immune-array analysis in sporadic inclusion body myositis reveals HLA-DRB1 amino acid heterogeneity across the myositis spectrum. Arthritis Rheumatol. 2017;69(5):1090-1099.
- [28] Ge Y, Lu X, Shu X, et al. Association of poly(ADP-ribose) polymerase 4 genetic variant rs188673928 with dermatomyositis/polymyositis in the Han Chinese population. Immunol Res. 2015;61(3):264-270.
- [29] Greenberg SA, Pinkus JL, Pinkus GS, et al. Interferon-α/β-mediated innate immune mechanisms in dermatomyositis. Ann Neurol. 2005;57(5):664-678.
- [30] Kissel JT, Mendell JR, Rammohan KW. Microvascular deposition of complement membrane attack complex in dermatomyositis. N Engl J Med. 1986;314(6):329-334.
- [31] Bruder J, Siewert K, Obermeier B, et al. Target specificity of an autoreactive pathogenic human γδ-T cell receptor in myositis. J Biol Chem. 2012;287(25):20986-20995.
- [32] Fasth AER, Dastmalchi M, Rahbar A, et al. T cell infiltrates in the muscles of patients with dermatomyositis and polymyositis are dominated by CD28null T cells. J Immunol. 2009;183(7):4792-4799.
- [33] Targoff IN. Myositis specific autoantibodies. Curr Rheumatol Rep. 2006;8(3):196-203.
- [34] Hengstman GJ, ter Laak HJ, Vree Egberts WT, et al. Anti-signal recognition particle autoantibodies: marker of a necrotising myopathy. Ann Rheum Dis. 2006;65(12):1635-1638.
- [35] Fiorentino DF, Chung LS, Christopher-Stine L, et al. Most patients with cancer-associated dermatomyositis have antibodies to nuclear matrix protein NXP-2 or transcription intermediary factor 1y. Arthritis Rheum. 2013;65(11):2954-2962.
- [36] Rider LG, Werth VP, Huber AM, et al. Measures of adult and juvenile dermatomyositis, polymyositis, and inclusion body myositis. Arthritis Care Res. 2011;63(S11):S118-S157.
- [37] Wang DX, Shu XM, Tian XL, et al. Intravenous immunoglobulin therapy in adult patients with polymyositis/dermatomyositis: a systematic literature review. Clin Rheumatol. 2012;31(5):801-806.
- [38] Degardin A, Morillon D, Lacour A, et al. Morphologic imaging in muscular dystrophies and inflammatory myopathies. Skeletal Radiol. 2010;39(12):1219-1227.
- [39] De Bleecker JL, De Paepe B, Aronica E, et al. 205th ENMC International Workshop: Pathology diagnosis of idiopathic inflammatory myopathies part II. Neuromuscul Disord. 2015;25(3):268-272.
- [40] Gordon PA, Winer JB, Hoogendijk JE, et al. Immunosuppressant and immunomodulatory treatment for dermatomyositis and polymyositis. Cochrane Database Syst Rev. 2012;(8):CD003643.

- [41] Moghadam-Kia S, Aggarwal R, Oddis CV. Treatment of inflammatory myopathy: emerging therapies and therapeutic targets. Expert Rev Clin Immunol. 2015;11(11):1265-1275.
- [42] Oddis CV, Reed AM, Aggarwal R, et al. Rituximab in the treatment of refractory adult and juvenile dermatomyositis and adult polymyositis: a randomized, placebo-phase trial. Arthritis Rheum. 2013;65(2):314-324.
- [43] Alexanderson H, Lundberg IE. Exercise as a therapeutic modality in patients with idiopathic inflammatory myopathies. Curr Opin Rheumatol. 2012;24(2):201-207.
- [44] Marie I, Hachulla E, Hatron PY, et al. Polymyositis and dermatomyositis: short term and longterm outcome, and predictive factors of prognosis. J Rheumatol. 2001;28(10):2230-2237.
- [45] Taborda AL, Azevedo P, Isenberg DA. Retrospective analysis of the outcome of patients with idiopathic inflammatory myopathy: a long-term follow-up study. Clin Exp Rheumatol. 2014;32(2):188-193.
- [46] Cox FM, Titulaer MJ, Sont JK, et al. A 12-year follow-up in sporadic inclusion body myositis: an end stage with major disabilities. Brain. 2011;134(Pt 11):3167-3175.